Nanopore Sequencing Technology and Tools: **Computational Analysis of the Current State, Bottlenecks and Future Directions** Damla Senol¹, Jeremie Kim^{1,3}, Saugata Ghose¹, Can Alkan² and Onur Mutlu^{3,1}

¹ Department of Electrical and Computer Engineering, Carnegie Mellon University, Pittsburgh, PA, USA ² Department of Computer Engineering, Bilkent University, Bilkent, Ankara, Turkey ³ Department of Computer Science, Systems Group, ETH Zürich, Zürich, Switzerland

Carnegie Mellon ETH zürich







Nanopore Sequencing

Nanopore Sequencing single molecule DNA sequencing o a **technology** that could potentially surpass current sequencing technologies o promises

Nanopore sequencers rely solely on the electrochemical structure of the different nucleotides for identification and measure the change in

Biological Nanopores for DNA Sequencing • first proposed in the 1990s, o recently made commercially available in May 2014 by Oxford Nanopore Technologies (ONT). MinION

- o higher throughput
- \circ lower cost
- o increased read length
- \circ no prior amplification step.
- It has one major drawback: **high error rates**.

the ionic current as long strands of DNA (ssDNA) pass *nano-scale* through the protein pores.

- o first commercial nanopore sequencing device
- high-throughput sequencing apparatus
- produces real-time data
- o inexpensive
- o pocket-sized / portable



Pipeline and Current Tools

Step 1. Basecalling

- Translates raw signal output of MinION to generate DNA sequences.
- Metrichor [1] (extracted with poretools [2]), nanonet [3], nanocall [4]

Step 2. Genome Assembly for noisy long reads

- o Using only the basecalled DNA reads, generates longer contiguous fragments called draft assemblies.
- o canu [5], miniasm [6]

Step 3. Polishing (Optional)

- Generates an improved consensus sequence from the draft assembly
- nanopolish [7], racon [8]

Our Goal

- o In order to take advantage of nanopore sequencing, it is important to increase the accuracy and the speed of the whole pipeline. o Although new nanopore chemistry R9 improves the data accuracy, the tools used for nanopore sequence analysis are of critical importance as they should overcome the high error rates of the technology.
- Our goal in this work is to comprehensively tools for nanopore analyze sequence analysis, with a focus on understanding the

[1] Metrichor. [https://nanoporetech.com/products/metrichor]

[2] Loman, Nicholas J., and Aaron R. Quinlan. "Poretools: a toolkit for analyzing nanopore sequence data." Bioinformatics 30.23 (2014): 3399-3401

[3] Nanonet. [https://github.com/nanoporetech/nanonet]

[4] David, Matei, et al. "Nanocall: An Open Source Basecaller for Oxford Nanopore Sequencing Data." bioRxiv (2016): 046086. [5] Berlin, Konstantin, et al. "Assembling large genomes with single-molecule sequencing and locality-sensitive hashing." Nature biotechnology 33.6 (2015): 623-630.

[6] Li, Heng. "Minimap and miniasm: fast mapping and de novo assembly for noisy long sequences." Bioinformatics (2016): btw152.

[7] Loman, Nicholas J., Joshua Quick, and Jared T. Simpson. "A complete bacterial genome assembled de novo using only nanopore sequencing data." Nature methods (2015).

[8] Vaser, Robert, et al. "Fast and accurate de novo genome assembly from long uncorrected reads." bioRxiv (2016): 068122.

Results

advantages, disadvantages, and bottlenecks of the various tools.







Assembled Genome

Mapping Assembled Contigs Against Reference Genome Basecaller: Metrichor, Assembler: Miniasm



Basecaller	Execution Time (Basecalling)	Assembler	Execution Time (Assembly)	# contigs	# bp	# bp / genome length	% Accuracy
Metrichor	-	Canu	2195m	1	4,586,878	0.9882	99.7769
		Miniasm	372 sec	1	4,477,194	0.9646	89.9817
Noporat	2517m	Canu	124m	2	4,559,230	0.9822	99.7809
manomet		Miniasm	60sec	6	3,549,677	0.9491	90.2123
Nanocall (Fast mode)	5359m	Canu	154m	0	-	-	-
		Miniasm	607sec	0	-	-	-

Mapping Assembled Contigs Against Reference Genome Basecaller: Nanonet, Assembler: Canu



• ONT's cloud-based basecaller, Metrichor and local basecaller, *nanonet* perform similarly with high accuracy. However, another local basecaller *nanocall* is **not suitable for R9 data**.

- o Canu, the assembler with error correction, produces highquality assemblies but is relatively slow compared to Miniasm, the assembler without error correction.

Mapping Assembled Contigs Against Reference Genome Basecaller: Nanonet, Assembler: Miniasm















